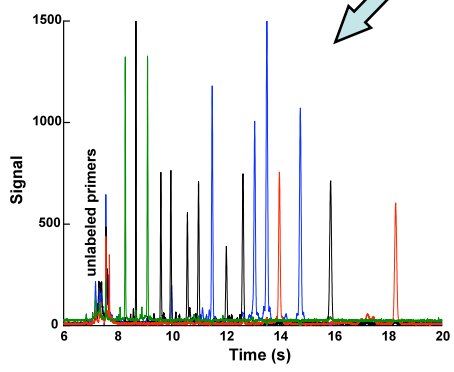
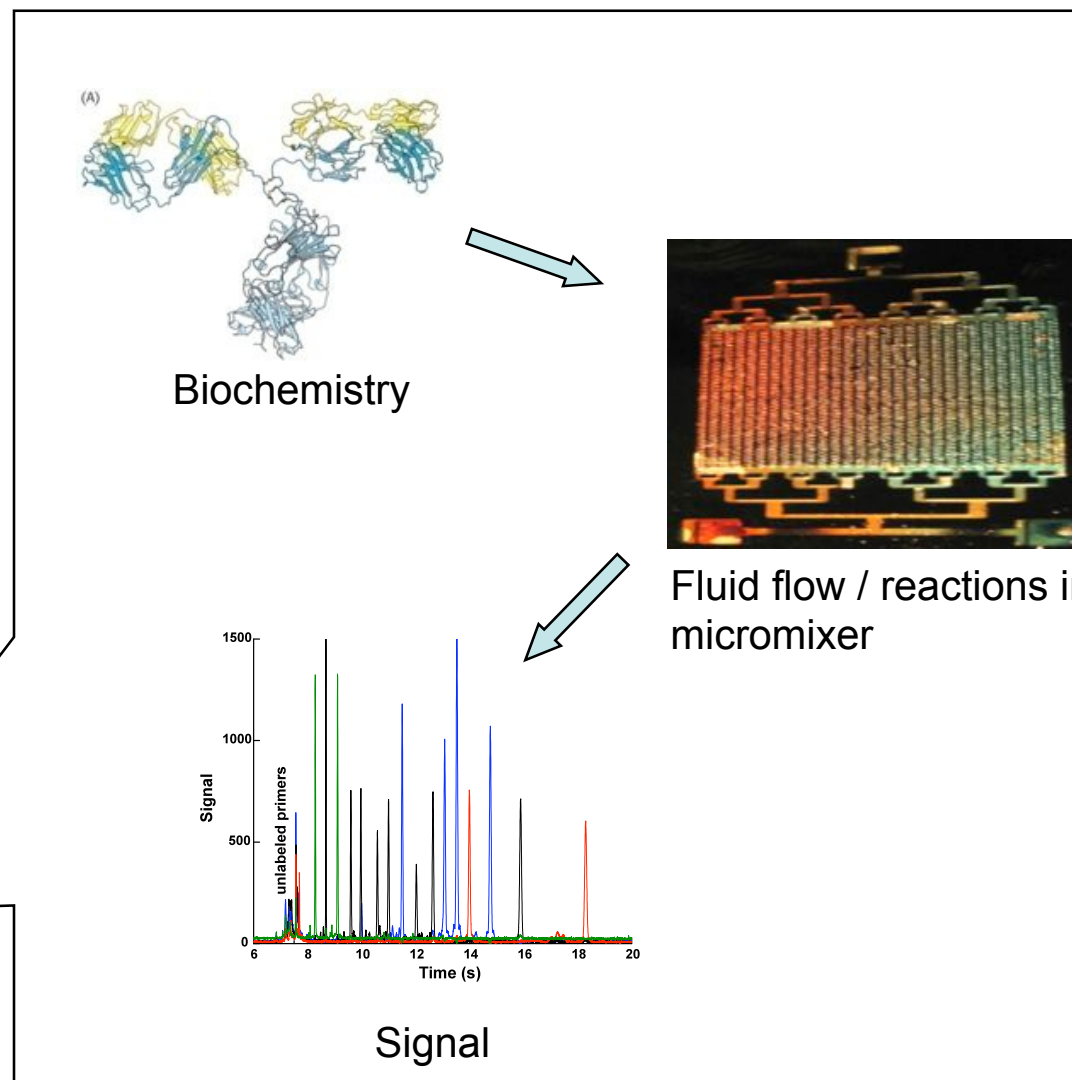
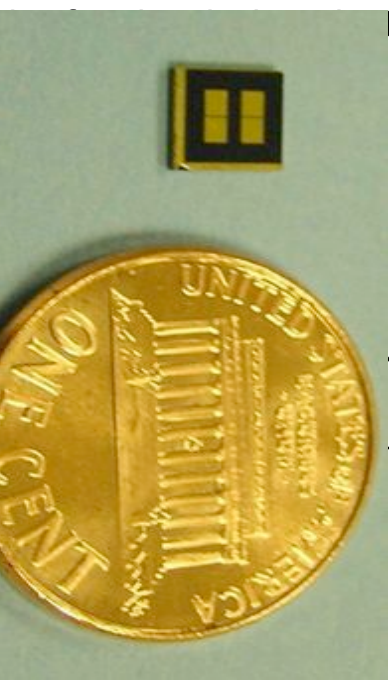


# Immunosensors

Ricardo Cortez, Mehnaaz Ali,  
Thomas Bishop, Kate Hamlington,  
Jerina Pillert, and Mangilal Agarwal

# Antibody-based Biosensor

System will be composed of fluidic and sensing elements (microfluidics) targeted for the detection of biological or chemical analytes. **Components:** microfluidic elements for sample processing, nanoporous membranes for target pre-concentration and carbon printed



# Antibody-based Biosensor

Tulane

LaTech IfM

Xavier

UNO

Merina Pillert  
Kate Hamlington  
Amit Jain  
Mehnaaz Ali  
Hank Ashbaugh  
Tom Bishop  
Diane Blake  
Ricardo Cortez  
Lisa Fauci  
Don Gaver

Senaka Kanakamedala  
Jie Liu  
Mangilal Agarwal  
Mark DeCoster  
Ji Fang  
Yuri Lvov

Robert Blake

Steven Rick

**Experiments:** characterization of antibodies, determination of assay parameters, preparation and reactivation of Apo-glucose oxidase, synthesis.

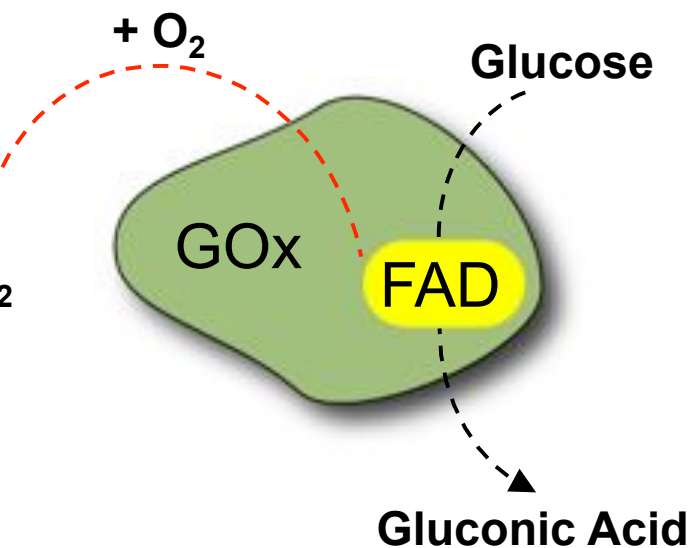
**MD Simulations:** antigens binding to antibody, energy minimization, loop structures, sequence alignment.

**CFD Simulations:** flows in microchannels, complex geometry, property optimization, reaction-diffusion-transport of concentrations, parallelization.

**Manufacturing:** microsensor layer fabrication, micromixer fabrication and evaluation, nanoporous membrane.

The immunosensor will use GOx mediated glucose oxidation for signal transduction

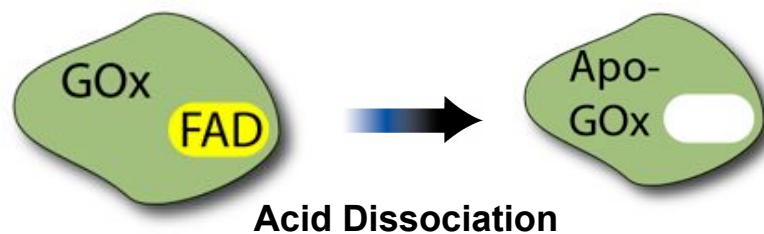
### Glucose Oxidation



Glucose oxidase requires the cofactor FAD for the catalysis of glucose to gluconic acid. This process involves the initial reduction of FAD to FADH<sub>2</sub> and subsequent oxidation by molecular O<sub>2</sub> generating H<sub>2</sub>O<sub>2</sub>

### E-Chem Sensor

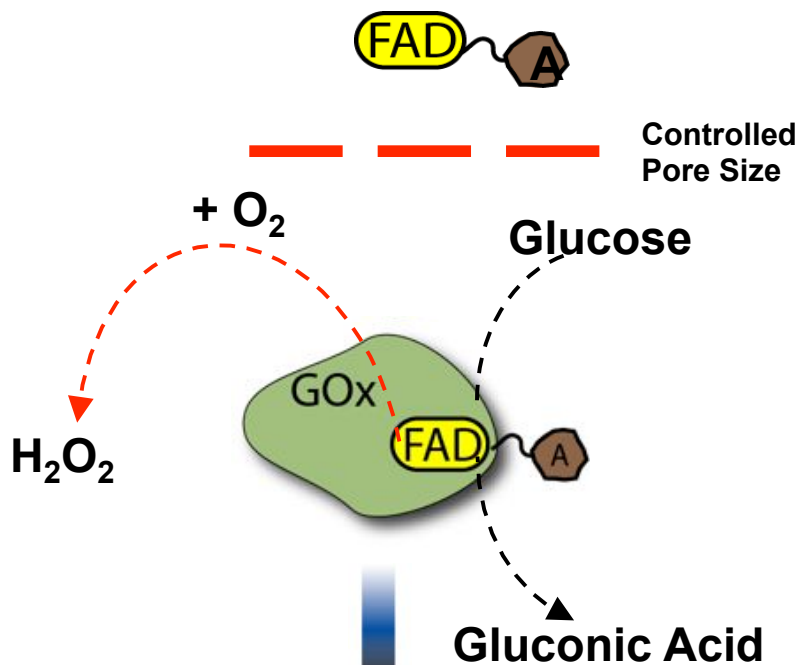
Enzyme activity can be modulated by the removal and introduction of the cofactor FAD. The cofactor can be efficiently dissociated under acidic conditions to yield apo glucose oxidase.



Thus the cofactor FAD can be conjugated to an analyte and utilized to modulate enzyme activity.

# General Strategy for E-chem Immunoassay

*Analyte conjugated FAD*



**FAD-analyte mediated reactivation of the Ap GOx in the presence of glucose.**

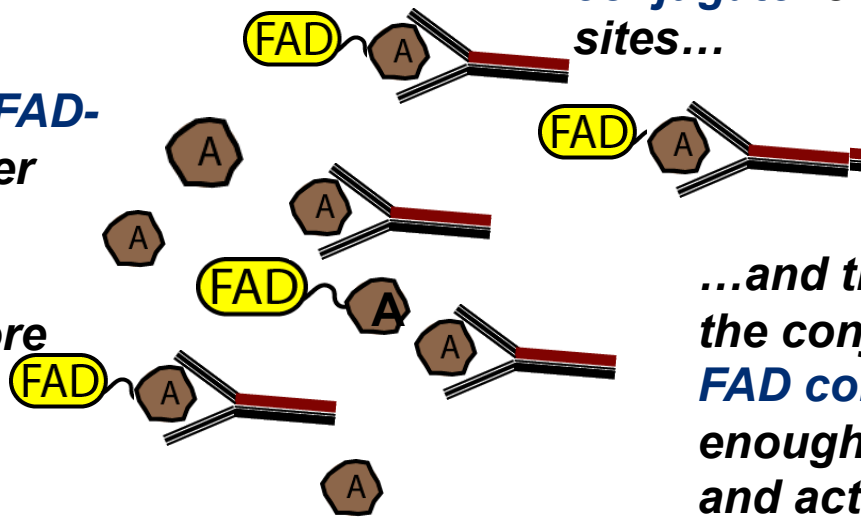
**-Glucose oxidase can be immobilized onto the carbon printed electrode (IFM, La Tech)**

**Electrochemical Sensor**

# E-Chemical Immunoassay

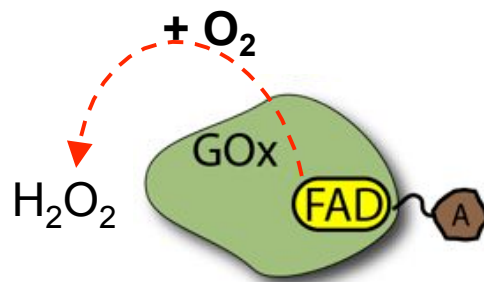
body is mixed with **FAD-analyte conjugate**. Larger body-analyte-FAD conjugate is not able to penetrate the control pore layer.

Addition of analyte from sample or environmental sample competes with the **FAD-analyte conjugate** for antibody binding sites...



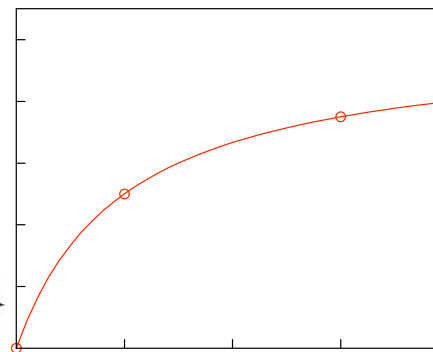
...and thus releases the conjugate. This **FAD conjugate** is small enough to enter pores and activate the enzyme.

Controlled Pore Size



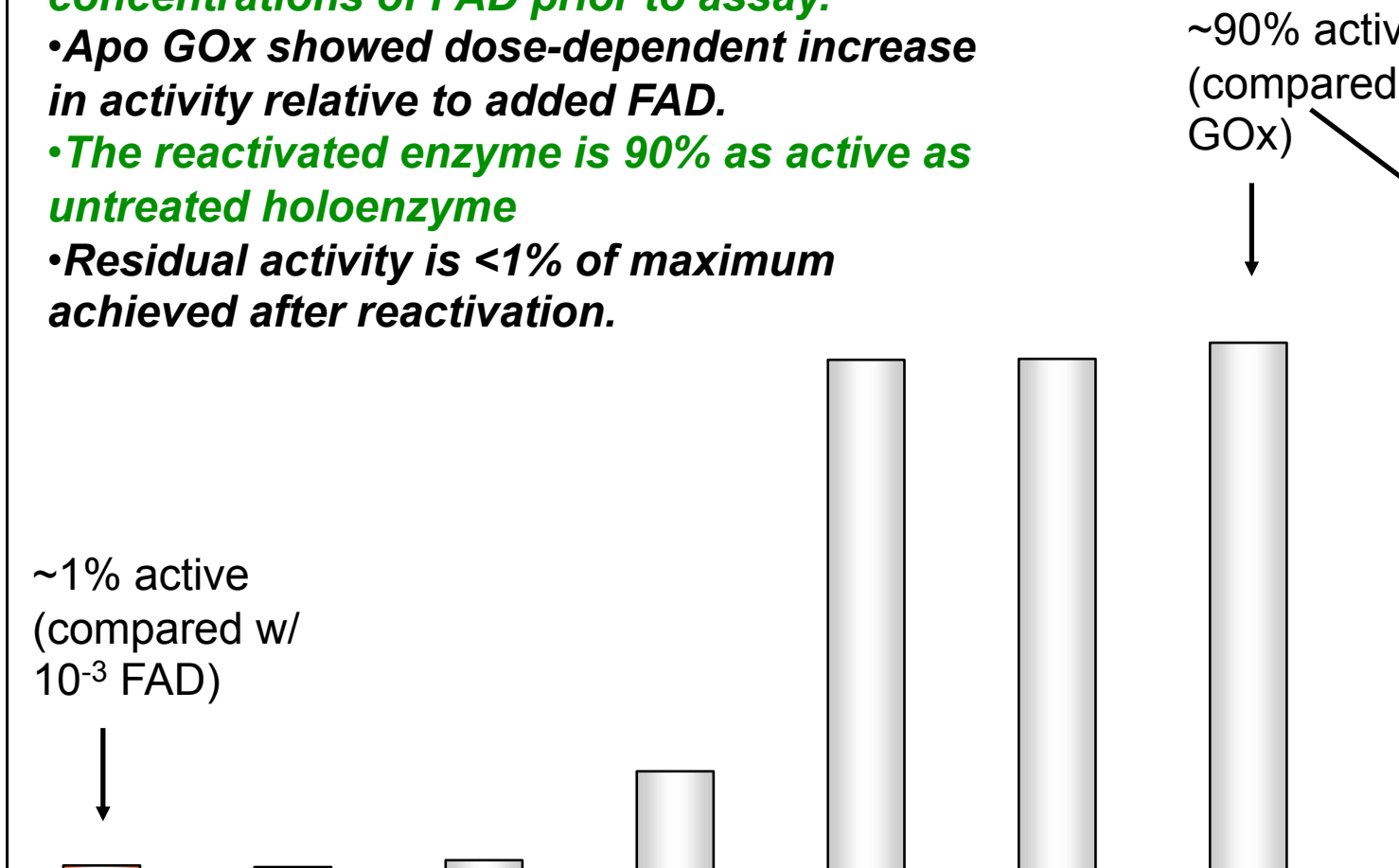
Signal

analyte concentration



# Reactivation of Apo-GOx

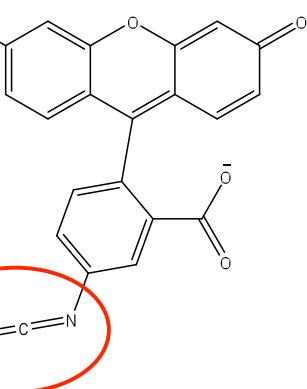
- **Apo GOx was incubated with increasing concentrations of FAD prior to assay.**
- **Apo GOx showed dose-dependent increase in activity relative to added FAD.**
- **The reactivated enzyme is 90% as active as untreated holoenzyme**
- **Residual activity is <1% of maximum achieved after reactivation.**



# Antibody – Analyte Selection

Antibody Number	Ligand	$K_d$ (M)	Availability
20	Fluorescein	$1.5 \times 10^{-9}$	Invitrogen
209	Fluorescein	$3.6 \times 10^{-9}$	Fitzgerald International
6	2,9-dicarboxyl-1,10 phenanthroline (DCP)	$7.5 \times 10^{-7}$	Blake et al., (2004) <i>Bioconj. Chem.</i> <b>15</b> :1125.
6	$UO_2^{2+}$ -DCP	$9.1 \times 10^{-10}$	Ibid
3	EDTA	$1.3 \times 10^{-8}$	Blake lab
3	$Cu^{2+}$ -EDTA	$2.2 \times 10^{-9}$	Blake lab

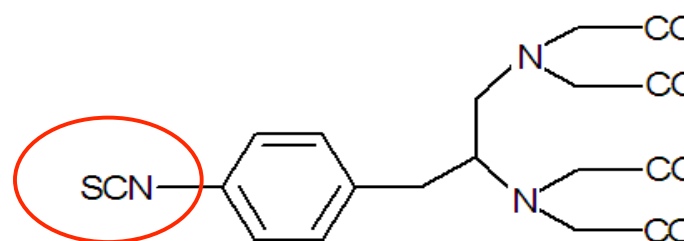
## Analytes to be coupled to FAD



**FITC**



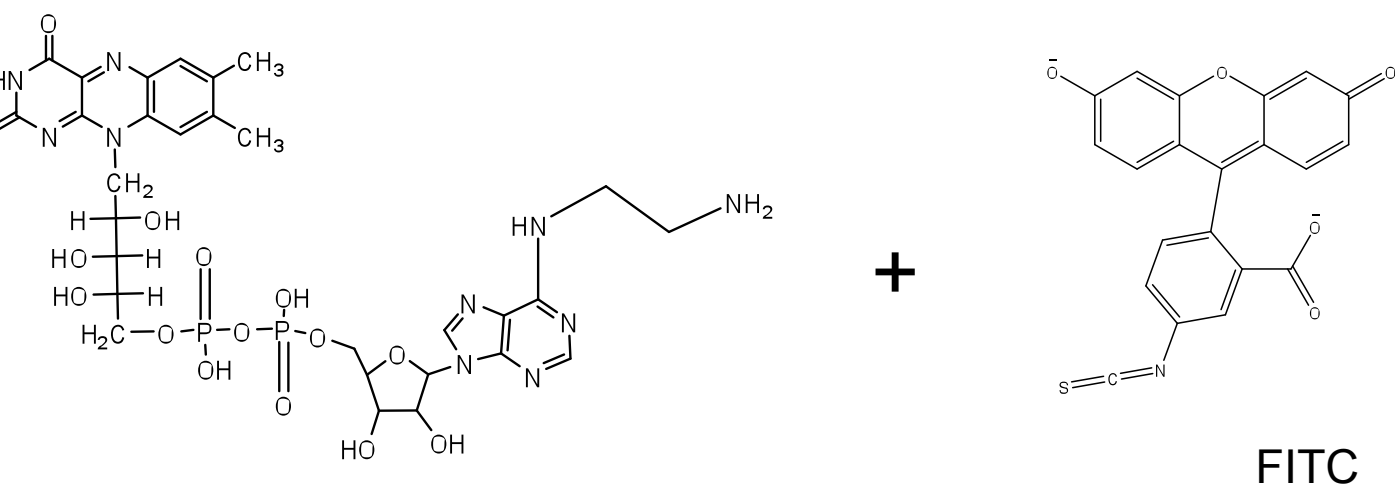
**DCP**



**EDTA**

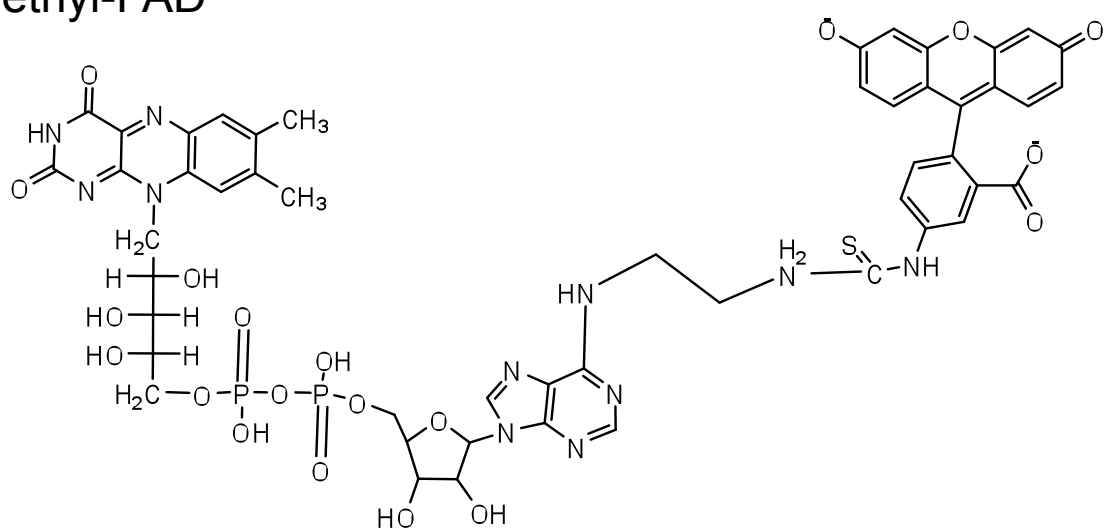


# Synthesis of FAD Conjugate



N<sup>6</sup>-2-aminoethyl-FAD

FITC



FAD-FITC conjugate

# Summary

## **Selection and Characterization of Antibodies**

*Commercial and in-house antibodies have been characterized.*

## **Apo-Glucose Oxidase has been prepared**

*Change in UV-VIS Spectra >300nm confirmed removal of FAD.  
Purification has been optimized to yield high quantity with low residual signal; storage conditions have been developed.  
Apo GOx has been transferred to LATech for sensor fabrication.*

## **FAD mediated Reactivation of Apo-Glucose Oxidase**

*Reactivation of Apo GOx was dependent on FAD concentration.  
Reactivated enzyme showed kinetics identical to native GO.  
Enzyme activity was not affected by components of the immunoassay.*

## **Synthesis of primary amine terminated FAD**

*N<sup>6</sup>-2-aminoethyl FAD has been synthesized and characterized.  
This intermediate was used to synthesize FAD-analyte conjugates.  
The apo enzyme could be reactivated with the FAD-FITC conjugate.  
A newly synthesized bifunctional crosslinker is also being tested for the preparation of FAD conjugates.*

# Antibody Bootcamp for Modelers

## Experimental Rotation in Blake Lab

Ashbaugh graduate student (Jain) spent one and half weeks in Blake lab learning experimental protocols for antibody sensing.

Titer experiments performed to measure concentrations of antibody 5B2,  $\text{Pb}^{2+}$ -DTPA-benzyl-BSA conjugate, metal chelator (DTPA), and  $\text{Pb}^{2+}$ -DTPA.

Enzyme-Linked ImmunoSorbant Assay (ELISA) used for titer of monoclonal antibody 5B2 and  $\text{Pb}^{2+}$  conjugate.

Competitive inhibition ELISA used to infer the ability of DTPA and  $\text{Pb}^{2+}$  to bind to 5B2.

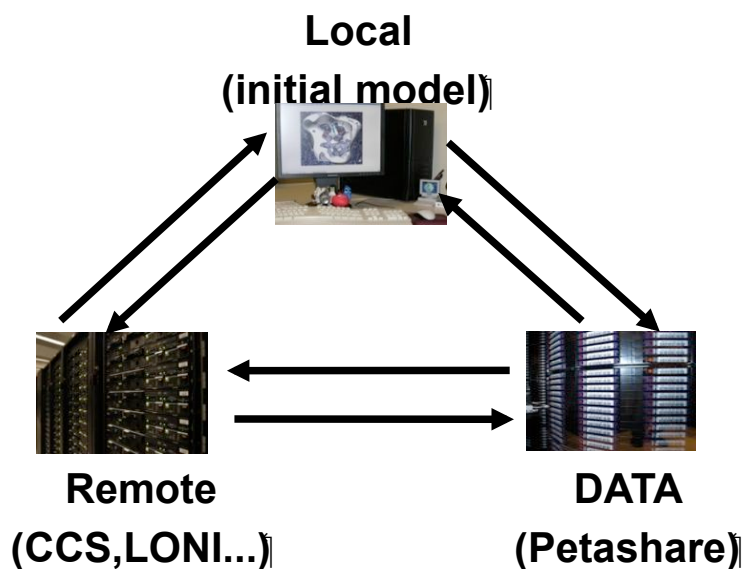
# The Molecular Modeling Requires

)Creation of putative antibody models based on sequence (Rosetta, I-TASSER, Modeller)

)Parameterization of the analytes that bind to the antibodies (AMBER, Gaussian)

)Docking analytes in different potential antibody binding sites (AutoDock, PackMol)

)Optimization of the antibody-analyte interaction by *in silico* point mutations (Methods under development, **REDS**)



# Sensors: Computational Aspects MD

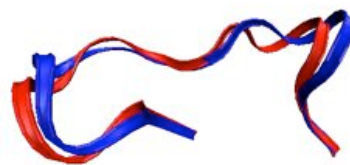
## Simulations of 5B2 loop region (Test Cases)

Binding of antigens to antibody occurs in loop domain. Aim to identify useful simulations side chains in loop region that contribute to binding specificity to guide antibody engineering.

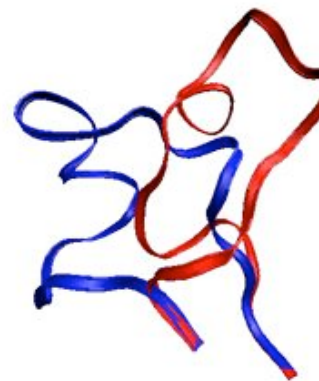
In vacuo energy minimizations of 5B2 LC and HC loops confirm previous identification of metal binding residue Lys<sup>58</sup>.

Replica Exchange Molecular Dynamic performed of 5B2 in vacuo and explicit solvent to generate families of loop structures for minimization to determine robustness of predictions and identify spatial and dynamic correlations between key binding residues

Initial findings: HC3 loop has more varied and flexible structure than the other antibody loops



**LC1**



**HC3**

# Sensors: Computational Aspects MD

## Simulations of 5B2 loop region (continued II)

### • Replica Exchange (REMD):

*replica*: several simultaneous simulations

2 levels of parallelization

*exchange*: simulations swap information

## Simulation Characteristics

### Loops

~1000 atoms => 2CPUS/sim

10ns run time => Gb's data;

Full REMD in 24hrs 64CPUS

### Full System

~10,000atoms => 4CPUS/sim

10ns run time => 10-100Gb data;

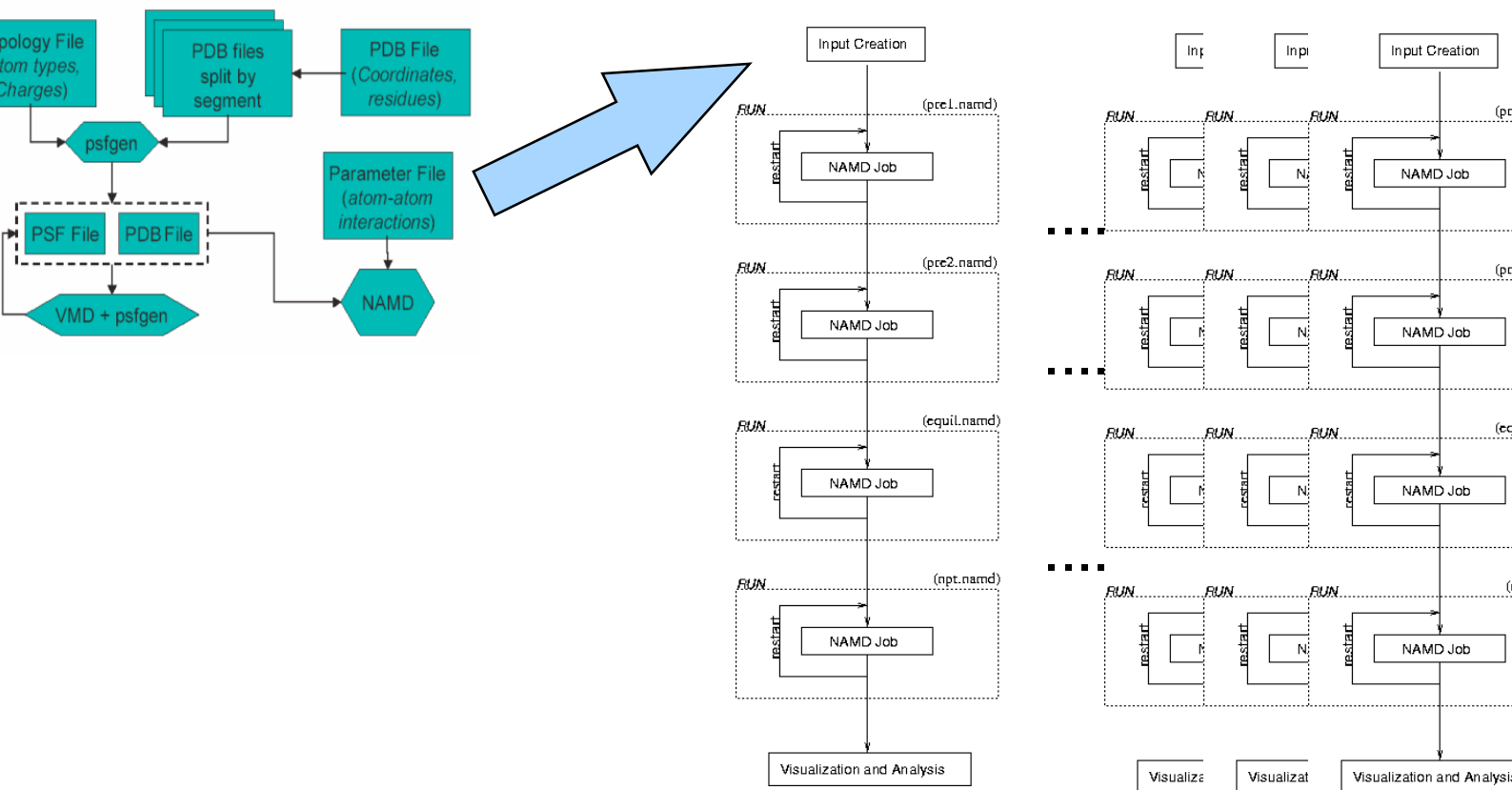
Full REMD in 2wks 64CPUS

# Biosensors: MD Fast Track Study

high throughput simulation workflow

Bishop (CCS @ TU)

Emir Embahsi & Tevik Kosar (CCT @ LSU)



# ***CyberTools* Connections**

## WP 1: Scheduling and Data Services.

The details of integrating our Molecular Modeling packages into WP 1 are being addressed by Drs. Thomas Bishop (Tulane) and Tevfik Kosar (LSU).

## WP 2: Information Services and Portals.

Drs. Thomas Bishop and Tevfik Kosar are collaborating to bring Bishop's A folding simulations on-line. The Workflow resulting from this effort can be readily modified to investigate the antibody and analyte interactions.

## WP 3: Visualization Services.

Work is in progress to create modules that will permit all scientists involved in the project to visualize molecular models and other results via a common web interface without the necessity of transferring data or installing software on local lab computers.

## WP 4: Application Services and Toolkits.

Drs. Steven Rick (UNO) and Henry Ashbaugh are developing replica simulation techniques that will enable this group to efficiently identify anti-peptide sequences that optimize the antibody-analyte interactions.



# Fluid Mechanics and Transport

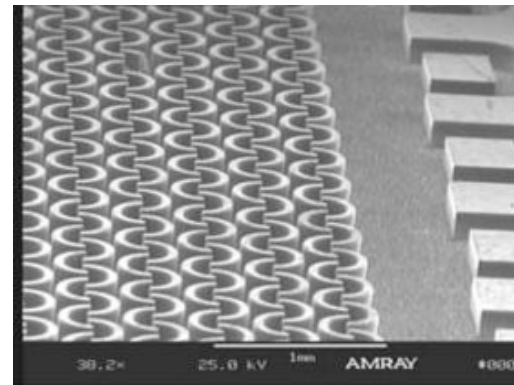
**GOAL** → *Computationally determine the optimal geometric configuration of the omega channel network to enhance mixing of two species.*

Laminar flow field governed by continuity & Stokes equations:

$$\nabla P = \mu \nabla^2 \mathbf{u}$$

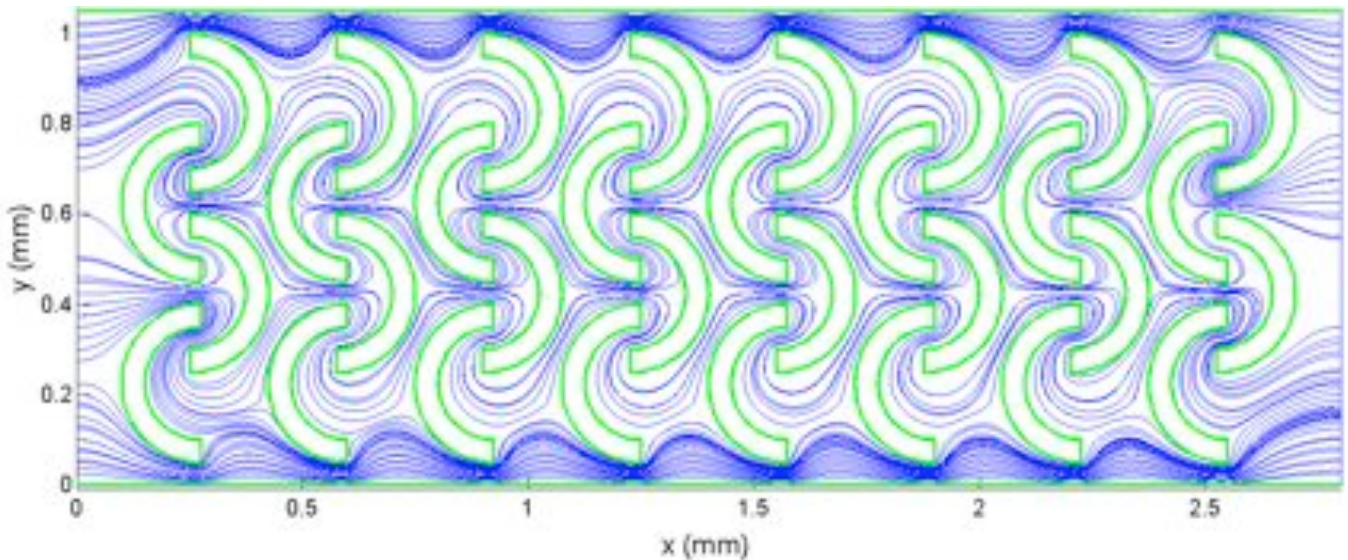
$$\nabla \cdot \mathbf{u} = 0$$

Boundary Element Method determines velocities and surface stresses

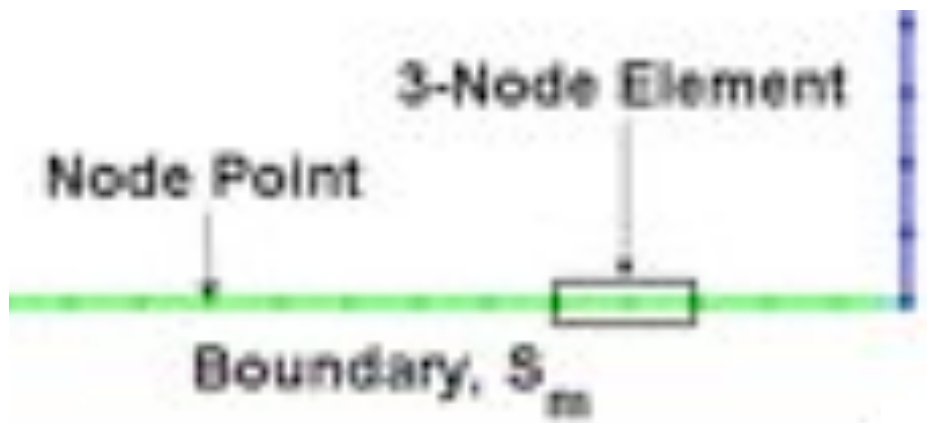


Omega channels developed by IfM

**Streamlines resulting from constant pressure drop across model channel**



# Boundary Element Method



Velocity  $\mathbf{u}$  and stress  $\boldsymbol{\tau}$  are approximated as quadratic polynomials, and at each node point, satisfy

$$\mathbf{C}_{ki}u_i(\mathbf{x}) + \sum_{m=1}^N \int_{S_m} \mathbf{T}_{ik}(\mathbf{x}, \mathbf{y})u_i(\mathbf{y})dS_m = \frac{1}{\mu} \sum_{m=1}^N \int_{S_m} \mathbf{U}_{ik}(\mathbf{x}, \mathbf{y})\tau_i(\mathbf{y})dS_m$$

Integral equation is expressed as system of linear equations:

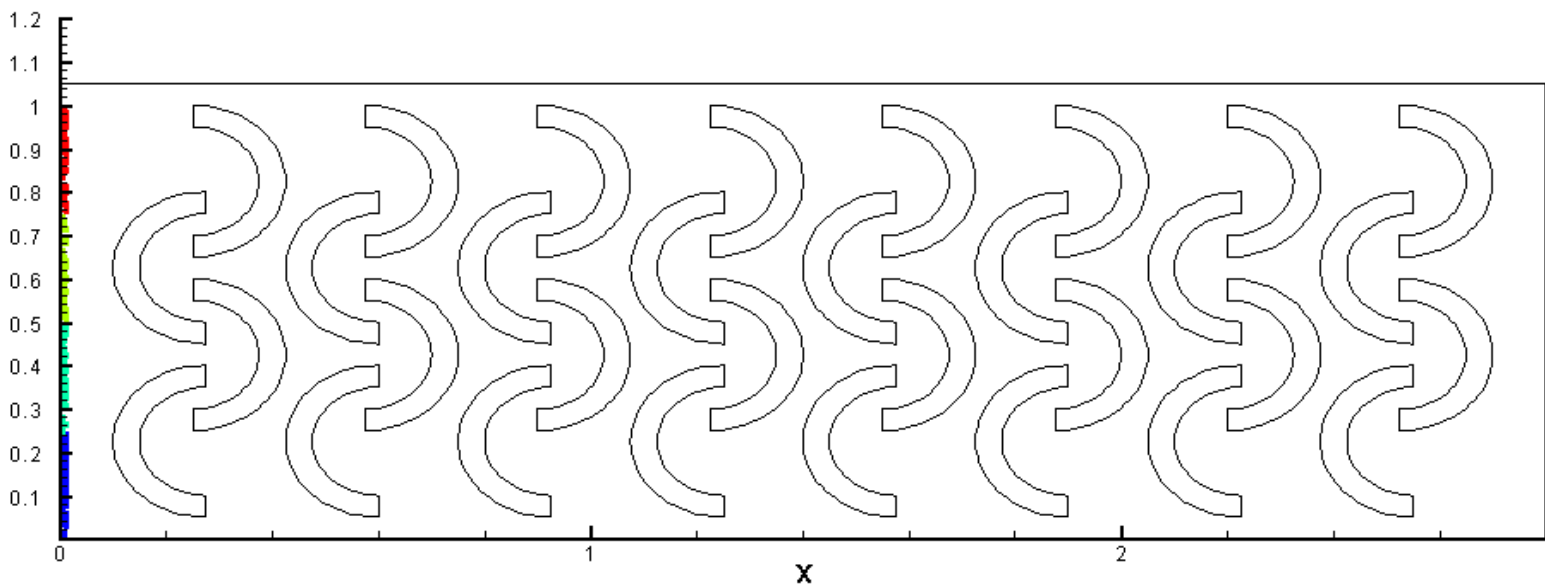
$$H\mathbf{u} = G\boldsymbol{\tau}$$

Elements of H and G computed using Gaussian quadrature rules

Optimization of simulation is being developed in conjunction with WP4 and will create a general purpose *CyberTool*.

# Results: Particle Trajectory

*For more info see our poster!*



Particles initially positioned along y-axis at  $x = 0$

Path of each particle traced as it flows through the domain

Note inner particles travel more slowly than outer particles that migrate quickly across channel along outer walls

Results suggest domain modification is important to improve mixing

# Microsensor Mixing and Transport

- Analyte-FAD conjugate and analyte from serum compete to bind with antibody
- Binding and release occur spontaneously as analytes and antibody are transported by fluid motion

**Each analyte/antibody satisfy a reaction-diffusion equation:**

$$\frac{\partial C_i}{\partial t} + \nabla \cdot (\mathbf{u} C_i) = D_i \nabla^2 C_i + R_i$$

$\mathbf{u}$  – fluid velocity

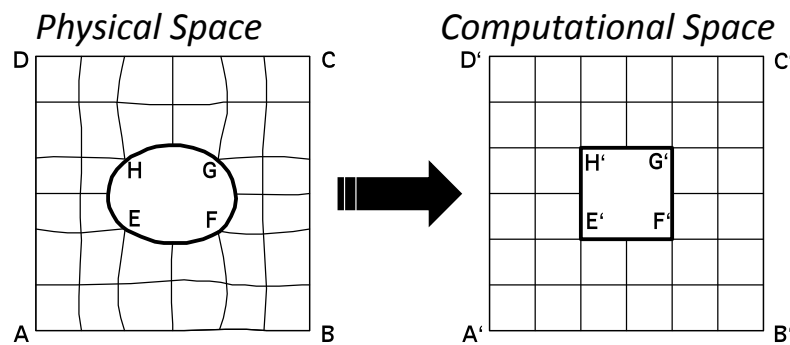
$C_i$  – concentration of each species

$D_i$  – diffusion coefficient

$R_i(C)$  – reaction term

# Transport Methodology

transform equations into a boundary-fitted coordinate system



use the Finite Volume Method to solve for *concentration*

Note: velocity field obtained from BEM code

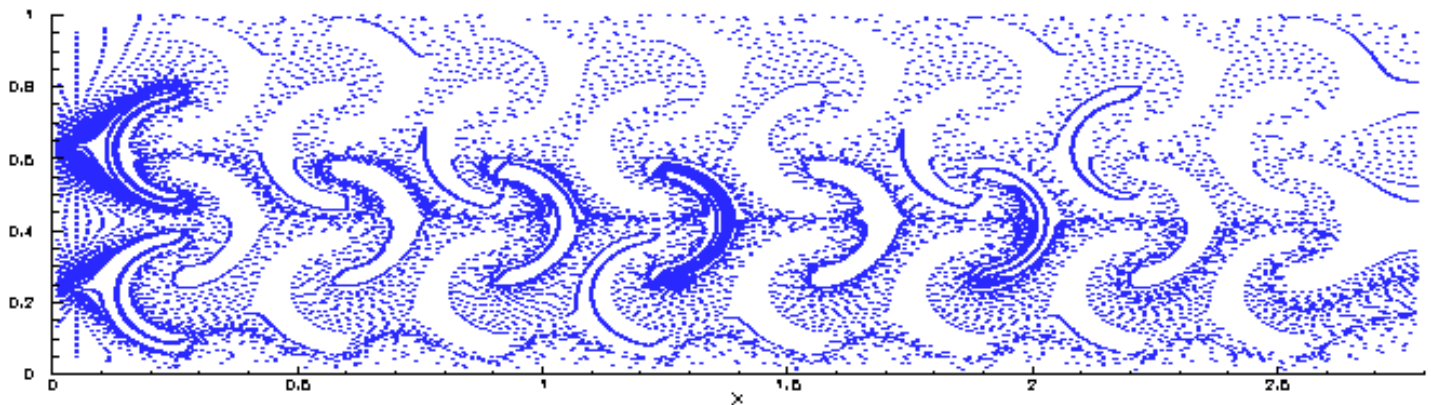
multiblock approach for omega channels

working with Dr. Blake for reaction/diffusion rates

# CyberTools Connections

## Current Work:

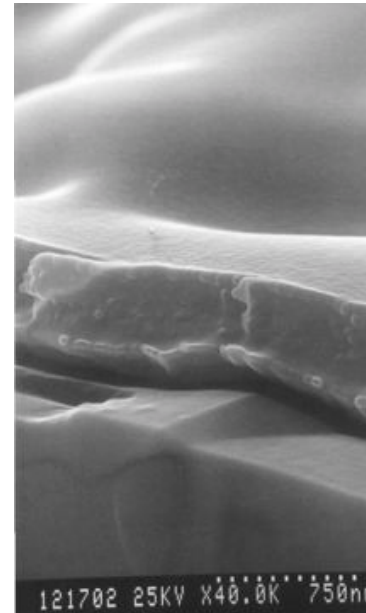
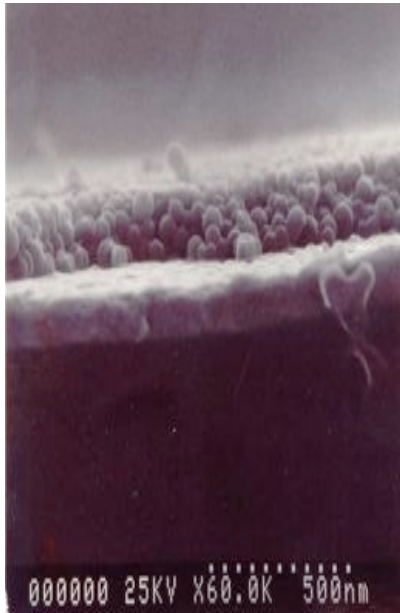
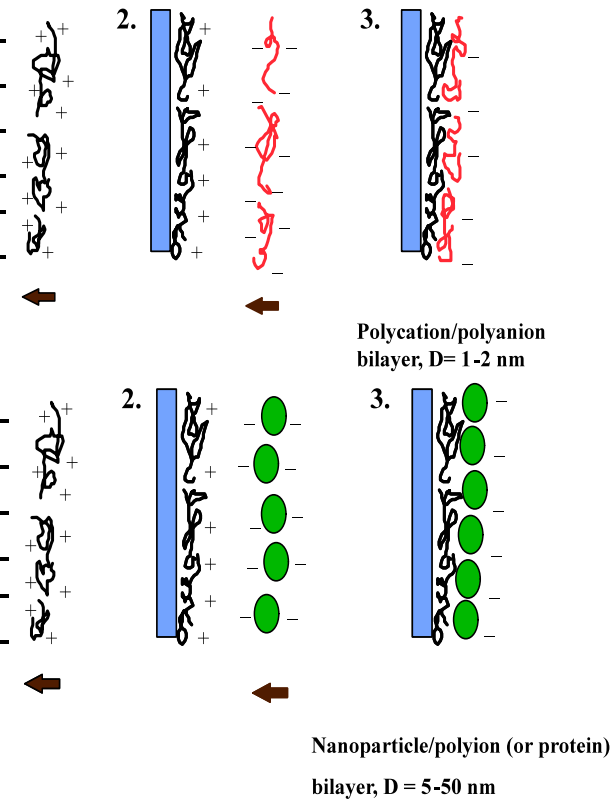
- Parallelization of Stokes flow problem for use in the HPC environment (WP4: Mayank Tyagi, Shantenu Jha, Sanjay Kodiyalam)
  - *OpenMP*
- Visualization of model problem using TecPlot (with WP3)
- Generalization of code to develop a *CyberTool* package that solves Stokes flow equations



## Future Work:

- Parallelization of source code including transport

# Layer-by-Layer (LbL) Nanoporous Membrane for Immunoassay (sensor technology for enzyme deposition)

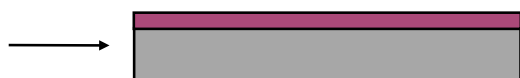


SEM cross-section images of (glucose oxidase/PAH)<sub>22</sub> multilayer on quartz (left) and (40 nm silica/PAH)<sub>6</sub> film on silver electrode (right).

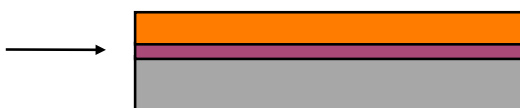
Principle of the layer-by-layer assembly by alternate adsorption of polycations and polyanions or nanoparticles

# Polymer-based Electronic Microsensor Fabrication

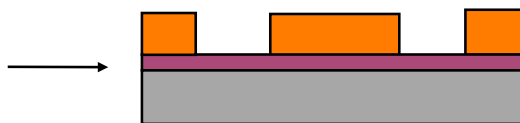
Silicon wafer with oxide layer



Spin coat PR 1813 resist layer



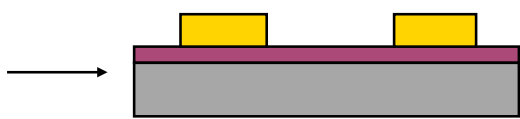
Pattern PR 1813 resist



Sputter gold electrodes



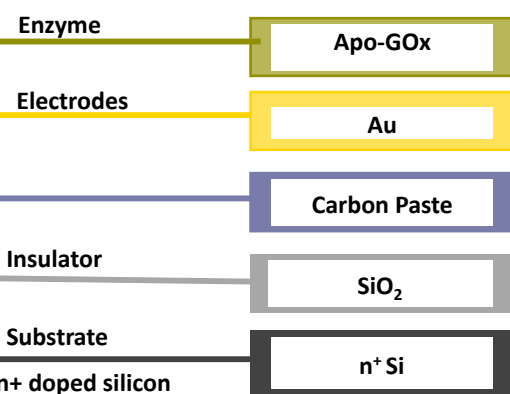
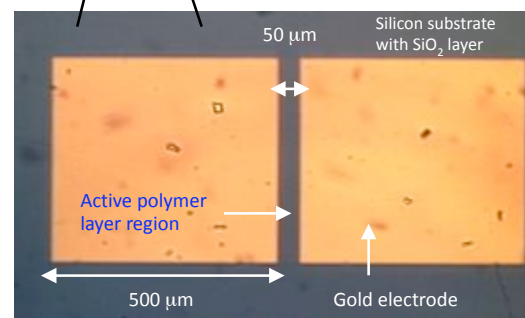
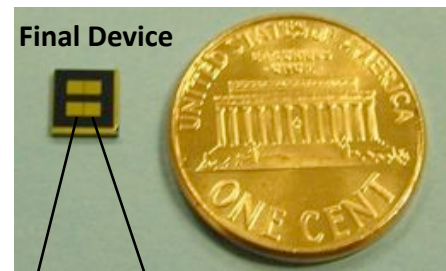
Lift-off PR 1813 resist



Final Device

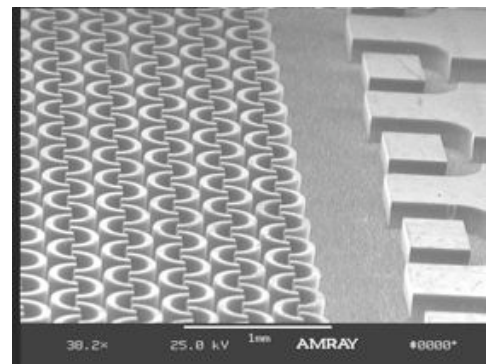
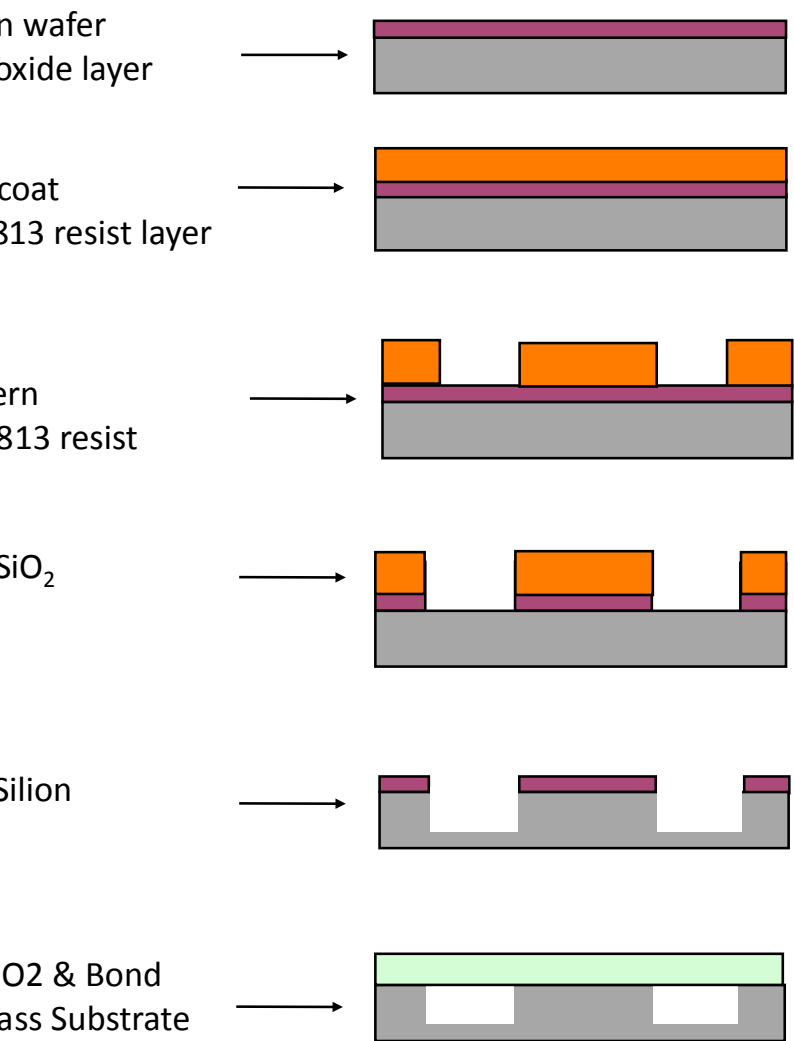


Final Device





# Micromixer Fabrication



SEM → Omega Channel Micromixer

## Fabrication

- Lithography
- ICP
- Bonding

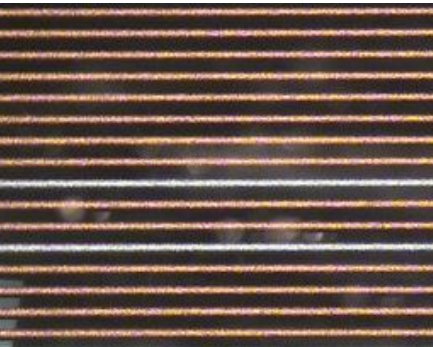
## Challenges

- Connectors

## Modifications

- New set of connectors from Upchurch Scientific are being tested and evaluated

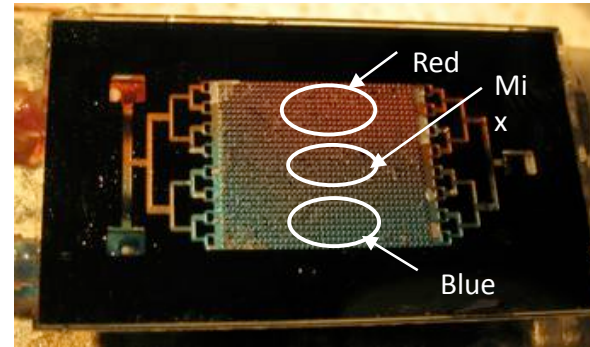
# Micromixer Evaluation



Straight Channel



Omega Channel



Micromixer

## Challenges

Laminar Flow

Mixing only at the center of the device

## Modifications

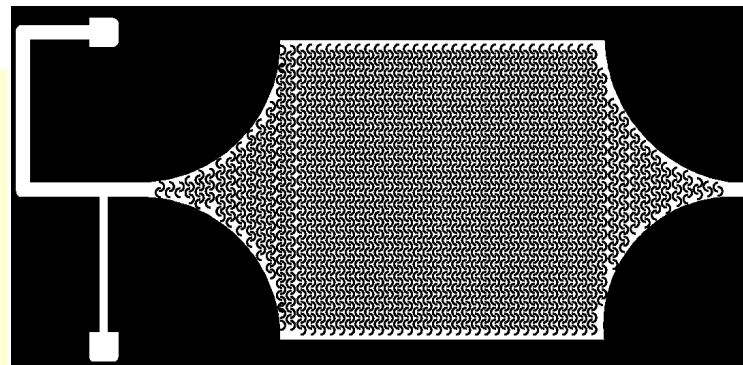
Designed 'T' shape inlet and outlet for

initial mixing

Quantification

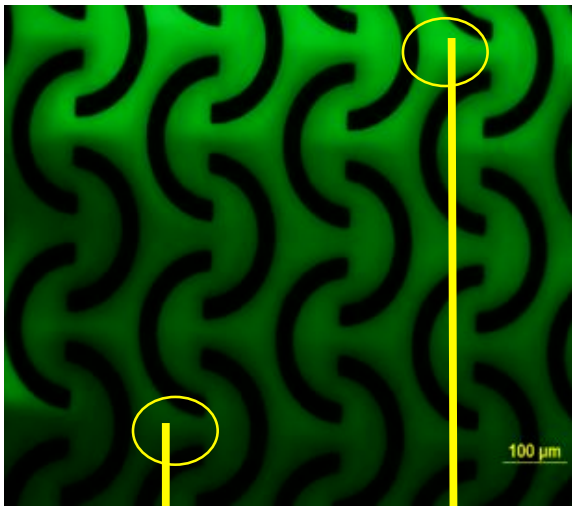
Image analysis software

Using fluorescent dyes for better signal/noise



New Micromixer Design

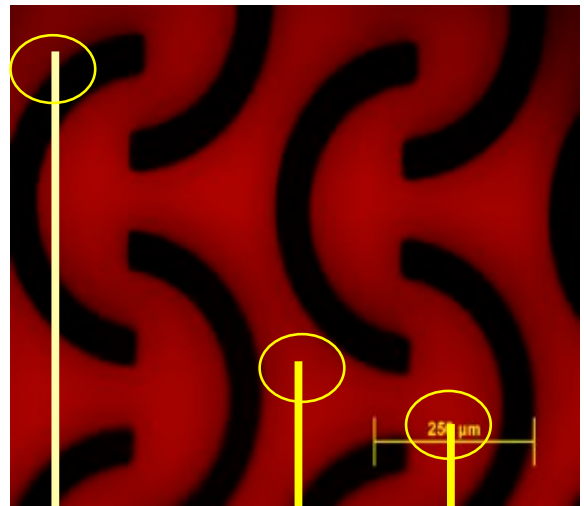
# Fluorescent dyes in the omega channels: VISUALIZED WITH MICROSCOPY AND DIGITAL CAMERA



Water

FITC

flow of fluids



Water

Rhodamine

Mixture of two

# Summary and Conclusions

## Microfluidic Component

- Fabricated and evaluated two sets of micromixers
- Designed new micromixer based on the results obtained (fabricated)

## Nanoporous Membrane

- LbL nanoassembly is being evaluated for fabricating nanoporous membrane; **New:** in collaboration with Dr. Scott Gold (Louisiana Tech.) for modeling of flow through porous membrane.

## Reproducibility

- Evaluating PEDOT and carbon nanotube based microsensor for reproducibility, Selectivity, and Life Time

## Testing Procedures

- Currently testing fluorescent dyes and particles (proposed) for evaluating micromixers.
- Currently evaluating microscale sensor system based on carbon nanotubes

- **Carbon-based Electrodes**

- Under testing and fabrication

- **CyberTools Connection**

- Access Grid (AG) video conference with Tulane (23 July 2008): simulations and experimental data. Evaluation of CyberTools link to visualization software: VisIt 1.9.1 (Windows version, DeCoster) thru the **Cactus Code** Link.

- VISIT OUR GRADUATE STUDENT POSTER! -- SENAKA KANAKAMEDALA-

# Final Remarks

**Interactions between the laboratory and the molecular dynamics groups**  
provided new models of protein structure that allow testing of hypotheses in silico  
prior to time-consuming laboratory experiments.  
Validated *in silico* predictions based on laboratory experiments.  
Identified methodological refinements based on laboratory results.

**Interactions between the laboratory and the micromanufacturing groups**  
broadened the kinds of hardware and electronics that can be used to construct the  
sensors.  
Prompted micromanufacturers to examine paradigms used to validate their devices  
(e.g. they have modified the molecular identity and concentration ranges of reagents  
used to test their microscale mixers).

**Interactions between the fluid mechanics and the micromanufacturing groups**  
provided data to determine boundary conditions used in the simulations.  
Established realistic geometries for fluid flow domains.  
Outlined a plan to reduce the number of fabrication trials needed to optimize  
the final device.